

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Breast carcinoma (NOS)	PHYSICIAN	ORDERING PHYSICIAN	SPECIMEN	SPECIMEN SITE
	NAME		MEDICAL FACILITY		SPECIMEN ID
	DATE OF BIRTH		ADDITIONAL RECIPIENT		SPECIMEN TYPE
	SEX		MEDICAL FACILITY ID		DATE OF COLLECTION
	MEDICAL RECORD #		PATHOLOGIST		SPECIMEN RECEIVED

Genomic Signatures

Microsatellite status - MS-Stable
Tumor Mutational Burden - 5 Muts/Mb

Gene Alterations

For a complete list of the genes assayed, please refer to the Appendix.

BRCA1 V1234fs*8

PTEN loss

BAP1 rearrangement intron 13

TP53 T256P

3 Disease relevant genes with no reportable alterations: **BRCA2, ERBB2, PIK3CA**

Report Highlights

- Targeted therapies with NCCN categories of evidence in this tumor type: **Olaparib** (p. 9), **Talazoparib** (p. 10)
- Variants that may inform nontargeted treatment approaches (e.g., chemotherapy) in this tumor type: **BRCA1 V1234fs*8** (p. 5)
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 12)
- Variants in select cancer susceptibility genes to consider for possible follow-up germline testing in the appropriate clinical context: **BRCA1 V1234fs*8** (p. 5)

GENOMIC SIGNATURES

Microsatellite status - MS-Stable

Tumor Mutational Burden - 5 Muts/Mb

GENE ALTERATIONS

BRCA1 - V1234fs*8

10 Trials see p. 12

PTEN - loss

10 Trials see p. 14

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Genomic Signatures section

No therapies or clinical trials. see Genomic Signatures section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
Olaparib <input type="checkbox"/>	Niraparib
Talazoparib <input type="checkbox"/>	Rucaparib
none	none

NCCN category

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING IN SELECT CANCER SUSCEPTIBILITY GENES

Findings below have been previously reported as pathogenic germline in the ClinVar genomic database and were detected at an allele frequency of >10%. See appendix for details.

BRCA1 - V1234fs*8 p. 5

This report does not indicate whether variants listed above are germline or somatic in this patient. In the appropriate clinical context, follow-up germline testing would be needed to determine whether a finding is germline or somatic.

GENE ALTERATIONS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Alterations section.

BAP1 - rearrangement intron 13 p. [7](#) **TP53 - T256P** p. [8](#)

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved through a centralized EU procedure or a national procedure in an EU Member State. Therapies, including but not limited to the following, have been approved nationally and may not be available in all EU Member States: Tretinoin, Anastrozole, Bicalutamide, Cyproterone, Exemestane, Flutamide, Goserelin, Letrozole, Leuprorelin, Triptorelin.

SAMPLE

ORDERED TEST #

GENOMIC SIGNATURES

GENOMIC SIGNATURE

Microsatellite status

RESULT
MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI is extremely rare in breast cancer, reported in 0-1% of cases across studies⁶⁻¹¹. The incidence of MSI is increased in triple-negative breast cancer⁹⁻¹¹ and in tumors with homologous recombination defects, such as mutations in BRCA1/2^{9,11}. Notably, in Lynch syndrome-related breast cancer, MSI has been reported in 51-85% of cases¹²⁻¹⁷. A prospective study of 123 patients with breast cancer treated with chemotherapy reported an increase in the incidence of MSI-H following chemotherapy treatment (from 0% pre-treatment to 19% post-treatment) and a significant association between MSI and tumor recurrence¹⁸.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁹. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁹⁻²¹. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers²²⁻²⁴. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{19,21,23-24}.



ORDERED TEST #

GENOMIC SIGNATURE

Tumor Mutational Burden

RESULT
5 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²⁵⁻²⁷, anti-PD-1 therapies²⁵⁻²⁸, and combination nivolumab and ipilimumab²⁹⁻³⁴. In multiple pan-tumor studies, increased tissue tumor mutational burden (TMB) was associated with sensitivity to immune checkpoint inhibitors^{25-28,35-39}. In the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors, significant improvement in ORR was observed for patients with TMB ≥ 10 Muts/Mb (as measured by this assay) compared with those with TMB < 10 Muts/Mb in a large cohort that included multiple tumor types³⁵; similar findings were observed in the KEYNOTE 028 and 012 trials²⁸. At the same TMB cutpoint, retrospective analysis of patients with solid tumors treated with any checkpoint inhibitor identified that tissue TMB scores ≥ 10 Muts/Mb were associated with prolonged time to treatment failure compared with scores < 10 muts/Mb (HR=0.68)³⁹. For patients with solid tumors treated with nivolumab plus ipilimumab in the CheckMate 848 trial, improved responses were observed in patients with a tissue TMB ≥ 10 Muts/Mb independent of

blood TMB at any cutpoint in matched samples⁴⁰. However, support for higher TMB thresholds and efficacy was observed in the prospective Phase 2 MyPathway trial of atezolizumab for patients with pan-solid tumors, where improved ORR and DCR was seen in patients with TMB ≥ 16 Muts/Mb than those with TMB ≥ 10 and < 16 Muts/Mb³⁸. Similarly, analyses across several solid tumor types reported that patients with higher TMB (defined as $\geq 16-20$ Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy compared with patients with higher TMB treated with chemotherapy⁴¹ or those with lower TMB treated with PD-1 or PD-L1-targeting agents²⁶.

FREQUENCY & PROGNOSIS

A study of 3,969 patients with breast cancer reported a median TMB of 2.63 mutations per megabase (Muts/Mb), with 5% of cases harboring TMB ≥ 10 Muts/Mb; median TMB was significantly higher in hormone receptor (HR)-negative and HER2-negative tumors than HR-positive or HER2-positive tumors⁴². The Breast Invasive Carcinoma TCGA analysis reported an average (non-silent) mutation load of 0.84 Muts/Mb for luminal A tumors, 1.38 Muts/Mb for luminal B tumors, 2.05 Muts/Mb for HER2-enriched tumors, and 1.68 Muts/Mb for basal-like tumors⁴³. In breast cancer, TMB is significantly higher in recurrent versus primary tumors, metastatic versus localized cancers, triple-negative versus HR-positive tumors, and CDH1-mutated versus CDH1-wildtype tumors^{42,44-45}. Among metastatic tumors, TMB-high samples have been reported more frequently in invasive lobular carcinoma (9-17% of cases, depending on the TMB cutoff to designate TMB-

high) than in invasive ductal carcinoma (2-8% of cases, depending on the cutoff), and TMB-high (at either cutoff) has not been observed in papillary carcinoma^{42,44-45}. Breast carcinoma harbors a median TMB of 3.8 muts/Mb, and 3.1% of cases have high TMB (> 20 muts/Mb)⁴⁶. In a large study of patients with breast cancer, hypermutation was more frequently observed in metastatic tumors than in primary tumors⁴². In a study of 14,867 patients with breast cancer, high TMB was associated with older age and metastatic disease but was not significantly associated with PD-L1 positivity using the TMB cutoff of ≥ 10 Muts/Mb⁴⁵. In estrogen receptor-positive breast cancer, increased TMB in tissue samples ($>$ mean of 1.25 Muts/Mb) associated with shorter OS (HR=2.02) in an analysis of the TCGA data⁴⁷.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁴⁸⁻⁴⁹ and cigarette smoke in lung cancer⁵⁰⁻⁵¹, treatment with temozolomide-based chemotherapy in glioma⁵²⁻⁵³, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁵⁴⁻⁵⁸, and microsatellite instability (MSI)^{54,57-58}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{26-27,35}.

ORDERED TEST #

GENE
BRCA1

ALTERATION

V1234fs*8

TRANSCRIPT ID

NM_007294

CODING SEQUENCE EFFECT

3700_3704delGTAA

VARIANT ALLELE FREQUENCY (% VAF)

59.3%

POTENTIAL TREATMENT STRATEGIES

– Targeted Therapies –

Alterations that inactivate BRCA1 or BRCA2 may confer sensitivity to PARP inhibitors⁵⁹⁻⁷⁶ or ATR inhibitors⁷⁷⁻⁷⁹. Clinical responses to PARP inhibitors have been reported for patients with either germline or somatic BRCA1/2 mutations^{60,65,68,75-76} and for patients with platinum-resistant or -refractory disease^{59,64,71,74}. The PARP inhibitors talazoparib and olaparib have shown significant clinical efficacy for patients with HER2-negative advanced breast cancer and a germline BRCA mutation in Phase 3 studies^{62,80}. In the Phase 3 BROCADE3 study for patients with HER2-negative breast cancer harboring deleterious germline BRCA mutations, the addition of veliparib to platinum chemotherapy, which was continued as a monotherapy if chemotherapy was discontinued, improved median PFS compared with placebo plus platinum chemotherapy (14.5 vs. 12.6 months; HR=0.71)⁸¹. In a Phase 1 trial of monotherapy treatment with the ATR inhibitor BAY1895344, 2 patients with deleterious BRCA1 alterations and platinum-refractory ovarian carcinoma experienced a PR or prolonged SD⁷⁷. In other Phase 1 trials of combination approaches, a patient with BRCA1-mutated ovarian carcinoma experienced prolonged SD from the ATR inhibitor berzosertib combined with topotecan⁷⁸; another patient with platinum- and PARP-inhibitory refractory ovarian cancer and an inactivating germline BRCA1 mutation experienced a PR from berzosertib plus

carboplatin⁸²; and a third patient with BRCA1-mutated triple-negative breast cancer (TNBC) experienced a PR to the ATR inhibitor ceralasertib combined with olaparib⁸³. Preclinical studies of BRCA1/2 inactivation in T-cell acute lymphoblastic leukemia (T-ALL)⁸⁴, ovarian carcinoma⁸⁵, and TNBC⁸⁶ showing reduced cell viability and increased DNA damage during ATR treatment further support the sensitivity of BRCA1-deficient cells to ATR inhibitors. The WEE1 inhibitor adavosertib has been evaluated as a monotherapy and in combination with PARP-inhibitor, olaparib. In a Phase 2 study for patients with PARP-resistant ovarian cancer, the combination of olaparib and adavosertib elicited improved clinical benefit (ORR: 29%; DCR: 89%) compared to adavosertib alone (ORR: 23%; DCR: 63%); however, in the BRCA-mutated cohort, no significant difference in clinical benefit was observed between the combination (ORR: 19%) and monotherapy (ORR: 20%) treatments⁸⁷. In a Phase 1 monotherapy trial of adavosertib that included 9 patients with BRCA1/2-mutated solid tumors, 2 patients with BRCA1-mutated cancers (1 with ovarian serous carcinoma and 1 with oral squamous cell carcinoma) achieved PRs, and a third patient with ovarian serous carcinoma harboring mutations in BRCA1 and TP53 experienced 14% tumor shrinkage prior to disease progression⁸⁸.

– Nontargeted Approaches –

Germline BRCA mutations are associated with benefit from platinum chemotherapeutic agents (NCCN Breast Cancer Guidelines, v2.2022). Inactivation of BRCA1 may also predict sensitivity to the DNA-damaging agents trabectedin and lurbinectedin⁸⁹⁻⁹⁸.

FREQUENCY & PROGNOSIS

In the Breast Invasive Carcinoma TCGA datasets, BRCA1 mutations have been reported in 2-4% of cases^{43,99}. A study of patients with sporadic breast cancer identified BRCA1 mutation in 9.3% (4/43) of cases¹⁰⁰. BRCA1 mutations account for approximately 4.6-7% of breast cancer cases in patients with a family history of breast

cancer¹⁰¹⁻¹⁰². A study reported decreased nuclear BRCA1 protein expression in breast carcinoma samples (n=22), as compared to normal breast tissue¹⁰³. For BRCA1 and BRCA2 mutation carriers, the risk of developing breast cancer by the age of 70 has been found to be approximately 57-65% and 39-49%, respectively, and a lifetime risk of up to 90% has also been reported¹⁰⁴⁻¹⁰⁶. One study reported that the presence of germline BRCA2 mutations was significantly associated with inferior PFS for patients with breast cancer treated with first-line CDK4/6 inhibitors plus endocrine combination therapy¹⁰⁷.

FINDING SUMMARY

The protein encoded by BRCA1 is involved in the maintenance of genomic stability, including DNA repair, cell cycle checkpoint, and chromosome segregation¹⁰⁸. Alterations such as seen here may disrupt BRCA1 function or expression¹⁰⁹⁻¹¹¹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the BRCA1 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with hereditary breast and ovarian cancer syndrome (ClinVar, Mar 2022)¹¹². Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Inactivating germline mutations in BRCA1 or BRCA2 are associated with autosomal dominant hereditary breast and ovarian cancer¹¹³⁻¹¹⁴, and the lifetime risk of breast and ovarian cancer in BRCA1/2 mutation carriers has been estimated to be as high as 87% and 44%, respectively¹¹⁵. Elevated risk for other cancer types, including gastric, pancreatic, prostate, and colorectal, has also been identified, with an increase in risk ranging from 20 to 60%¹¹⁶. The estimated prevalence of deleterious germline BRCA1/2 mutations in the general population is between 1:400 and 1:800, with an approximately 10-fold higher prevalence in the Ashkenazi Jewish population^{115,117-122}. In the appropriate clinical context, germline testing of BRCA1 is recommended.

ORDERED TEST #

GENOMIC FINDINGS

GENE
PTEN

ALTERATION
loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

PTEN loss or mutation leads to activation of the PI3K-AKT-mTOR pathway and may predict sensitivity to inhibitors of this pathway¹²³⁻¹²⁶. Patients with PTEN-mutated HR+ HER2- breast cancer benefited from the addition of fulvestrant to the AKT inhibitor capivasertib in a Phase 2 study (mPFS 9.5 vs. 5.2 months, not significant), and patients who were HR+ experienced an ORR of 21% (for those exposed to fulvestrant) and 8% (for those who were fulvestrant-naive) on the combination in a Phase 1 trial¹²⁷⁻¹²⁸. In a Phase 2 study of capivasertib monotherapy for patients with PTEN- or AKT1-mutated breast cancer (ER+ or ER-), the ORR was 18% (2/11)¹²⁹. The Phase 3 IPATunity130 trial for patients with AKT1-, PTEN-, or PIK3CA-mutated HR+ HER2- breast cancer reported no significant PFS improvement with the first-line AKT inhibitor ipatasertib in combination with paclitaxel relative to paclitaxel alone (9.3 vs. 9.3 months)¹³⁰, despite Phase 1 results showing a potential benefit¹³¹. In a Phase 2 study of the PIK3CA inhibitor buparlisib for patients with PTEN-negative HR+ HER2- breast cancer, the mPFS was 2.5 months (n=3)¹³². Clinical studies in breast cancer have not observed an association between PTEN deficiency and response to the mTOR inhibitors everolimus or temsirolimus¹³³⁻¹³⁸, although exploratory analysis of Phase 3 studies suggests that patients with HER2+ metastatic breast cancer and PTEN loss derived significant benefit from everolimus added to trastuzumab plus chemotherapy¹³⁹. One PR has been observed in a patient with breast cancer

harboring PTEN and STK11 alterations following treatment with the mTORC1/2 inhibitor sapanisertib combined with metformin¹⁴⁰. A Phase 2 trial of the ATKi inhibitor capivasertib with paclitaxel versus paclitaxel alone showed a median OS benefit (19.1 vs. 13.5 months) both for patients with AKT1-, PTEN-, or PIK3CA-mutated triple-negative breast cancer (TNBC) (HR=0.58, 95% CI 0.2-1.6) and for patients with TNBC without PI3K-pathway mutations (HR=0.74, 95% CI 0.47-1.18)¹⁴¹. Despite promising initial results in earlier trials, the Phase 3 IPATunity130 trial for patients with AKT1-, PTEN-, or PIK3CA-mutated TNBC failed to show improved PFS for the first-line AKT inhibitor ipatasertib in combination with paclitaxel relative to paclitaxel alone (7.4 vs. 6.1 months)¹⁴²⁻¹⁴³. Preclinical data indicate that PTEN loss or inactivation may predict sensitivity to PARP inhibitors¹⁴⁴⁻¹⁴⁸, and clinical benefit has been observed for patients with PTEN-altered breast cancer including triple negative breast cancer¹⁴⁹, ovarian cancer¹⁵⁰, uterine leiomyosarcoma¹⁵¹, and endometrial cancer¹⁴⁸ treated with PARP inhibitors. However, some studies have reported a lack of association between PTEN mutation and PARP inhibitor sensitivity^{64,152}.

— Potential Resistance —

A multivariate analysis showed that patients with metastatic triple negative breast cancer (TNBC) harboring PTEN alterations were more likely to progress and experience shorter PFS and OS when treated with anti-PD-1/L1 therapies compared with patients with wildtype PTEN¹⁵³. Clinical and preclinical evidence suggests that PTEN loss or mutation may predict resistance to PI3K inhibitors¹⁵⁴⁻¹⁵⁶, and to CDK inhibitors such as palbociclib, ribociclib, and abemaciclib^{154,157}.

FREQUENCY & PROGNOSIS

In the TCGA dataset, PTEN mutation has been

reported in 4% of breast invasive carcinomas, while putative homozygous deletion of PTEN has been reported in 2% of cases⁴³. PTEN mutation has also been observed in 5.3% (1/19) of metaplastic breast cancers¹⁵⁸ and 2% of invasive lobular carcinoma tumors analyzed¹⁵⁹. PTEN mutations are associated more frequently with triple-negative breast cancer than with HER2- or hormone-positive breast cancer¹⁶⁰⁻¹⁶¹. Loss or reduction of PTEN expression has been observed in 28% of invasive ductal carcinomas and has been correlated with metastasis and poor patient prognosis, including decreased 2-year disease-free survival¹⁶²⁻¹⁶⁴.

FINDING SUMMARY

PTEN encodes an inositol phosphatase that functions as a tumor suppressor by negatively regulating the PI3K-AKT-mTOR pathway; loss of PTEN can lead to uncontrolled cell growth and suppression of apoptosis¹²⁴. Alterations such as seen here may disrupt PTEN function or expression¹⁶⁵⁻²⁰⁶.

POTENTIAL GERMLINE IMPLICATIONS

PTEN mutations underlie several inherited disorders, collectively termed PTEN hamartoma tumor syndrome (PHTS), which include Cowden syndrome (CS) and its variant Lhermitte-Duclos disease (LD), Bannayan-Riley-Ruvalcaba syndrome (BRRS), PTEN-related Proteus syndrome (PS), and Proteus-like syndrome²⁰⁷⁻²⁰⁸. The mutation rate for PTEN in these disorders ranges from 20 to 85% of patients^{207,209}. The estimated incidence of Cowden syndrome is 1/200,000, which may be an underestimate due to the high variability of this disorder²⁰⁷. Given the association between PTEN and these inherited syndromes, in the appropriate clinical context, germline testing for mutations affecting PTEN is recommended.

ORDERED TEST #

GENE

BAP1

ALTERATION

rearrangement intron 13

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Clinical²¹⁰ and preclinical²¹¹ evidence in the context of mesothelioma suggests that tumors with BAP1 inactivation may be sensitive to EZH2 inhibitors such as tazemetostat. Preclinical studies suggest that BAP1 is involved in the DNA damage response²¹²⁻²¹⁵, and BAP1 inactivation might be associated with sensitivity to PARP inhibitors²¹³⁻²¹⁴. One preclinical study suggests that HDAC inhibitors may be beneficial in BAP1-mutated uveal melanoma; however, it is unclear if these inhibitors are effective in other BAP1-mutated cancers²¹⁶.

— Potential Resistance —

One preclinical study suggests that BAP1 inactivation in breast cancer may be associated with resistance to tamoxifen²¹⁷.

FREQUENCY & PROGNOSIS

BAP1 somatic mutations are reported to be rare in breast cancer²¹⁸, and have been reported in 0.4-2% of breast invasive carcinoma cases^{43,99,219-220}. BAP1 has been suggested to play a tumor suppressive role in breast cancer cells by regulating genomic stability; high BAP1 expression was significantly associated with improved overall survival in patients with breast cancer, and with prolonged progression-free survival in patients with basal or luminal breast cancer²²¹.

FINDING SUMMARY

BAP1 (BRCA1 associated protein-1) encodes a ubiquitin hydrolase, a protein involved in regulating the availability of target proteins for the ubiquitin-proteasome protein degradation pathway; BAP1 is located on chromosome 3p21.3, in a region of frequent loss of heterozygosity

(LOH) in breast and lung cancer, and has been postulated to be a tumor suppressor²²²⁻²²³. Alterations such as seen here may disrupt BAP1 function or expression²²³⁻²³².

POTENTIAL GERMLINE IMPLICATIONS

BAP1 germline inactivating alterations, including mutations and deletions, are associated with BAP1 tumor predisposition syndrome (BAP1-TPDS), an autosomal-dominant syndrome characterized by early onset of benign melanocytic skin tumors^{226,233-234}. An estimated 2% of patients with BAP1-inactivated melanocytic tumors display germline BAP1 mutations²³⁵. Later in life, patients have an increased risk of cancers such as uveal melanoma, mesothelioma, clear cell renal cell carcinoma, basal cell carcinoma, and meningioma^{225-229,236}. In small studies, the prevalence of pathogenic germline BAP1 mutation has been reported as 22% in familial uveal melanoma and 4.4% in mesothelioma²³⁷⁻²³⁸. In the appropriate clinical context, germline testing of BAP1 is recommended.

SAMPLE

ORDERED TEST #

GENOMIC FINDINGS

GENE

TP53

ALTERATION

T256P

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

766A>C

VARIANT ALLELE FREQUENCY (% VAF)

29.6%

POTENTIAL TREATMENT STRATEGIES

– Targeted Therapies –

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib²³⁹⁻²⁴², or p53 gene therapy and immunotherapeutics such as SGT-53²⁴³⁻²⁴⁷ and ALT-801²⁴⁸. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype²⁴⁹. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer²⁵⁰. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer²⁵¹. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone²⁵². In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with

adavosertib combined with paclitaxel²⁵³. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53 alterations²⁵⁴. The Phase 2 FOCUS4-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring²⁵⁵. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage²⁴⁷. Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246²⁵⁶⁻²⁵⁸. In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR²⁵⁹. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies²⁶⁰⁻²⁶¹; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies²⁶²⁻²⁶³. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in breast cancer; mutations in this gene have been identified in 27-37% of breast carcinoma samples^{43,219,264-267}. TP53 mutations that are located within the region encoding the DNA binding domain are associated with poor prognosis in patients with breast cancer^{265,268-269}. TP53 mutation is also implicated in breast cancer susceptibility, as TP53 mutation carriers have an 18-60 fold increased risk for early onset breast

cancer²⁷⁰⁻²⁷².

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers²⁷³. Alterations such as seen here may disrupt TP53 function or expression²⁷⁴⁻²⁷⁸.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers²⁷⁹⁻²⁸¹, including sarcomas²⁸²⁻²⁸³. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000²⁸⁴ to 1:20,000²⁸³. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30²⁸⁵. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion²⁸⁶⁻²⁹¹. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy²⁸⁶⁻²⁸⁷. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease²⁹². Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{290,293-294}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

Olaparib

Assay findings association

BRCA1
V1234fs*8

AREAS OF THERAPEUTIC USE

The PARP inhibitor olaparib is available in the EU as a monotherapy to treat patients with BRCA-mutated high-grade epithelial ovarian, Fallopian tube, or peritoneal cancer. It is also available in combination with bevacizumab for patients with HRD-positive high-grade epithelial ovarian, Fallopian tube, or peritoneal cancer. It is additionally available as a monotherapy for patients with HER2-negative germline BRCA-mutated breast cancer, BRCA-mutated castration-resistant prostate cancer, and germline BRCA-mutated pancreatic adenocarcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of extensive clinical evidence in ovarian cancer⁶⁹⁻⁷³ as well as strong clinical evidence in multiple other cancer types^{59-61,69,72,76,295}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to olaparib.

SUPPORTING DATA

A Phase 3 study of olaparib monotherapy for patients with germline BRCA1/2 (gBRCA1/2)-mutated HER2-metastatic breast cancer reported a significantly longer median PFS (7.0 vs. 4.2 months, HR=0.58) and a higher ORR (60% vs. 29%) compared with standard chemotherapy⁶². The Phase 3 OlympiA trial of adjuvant olaparib for patients with gBRCA1/2-mutated breast cancer reported significantly increased invasive disease-free survival (IDFS) (HR=0.63), distant DFS (DDFS) (HR=0.61), and OS (HR=0.68; p=0.0091) compared with placebo; 4-year rates of IDFS, DDFS, and OS comparing

olaparib with placebo were 83% versus 75%, 87% versus 79%, and 90% versus 86%, respectively²⁹⁶. Phase 2 studies of olaparib monotherapy for patients with BRCA-mutated advanced breast cancer reported median PFS of 3.7 to 5.7 months and high clinical benefit rates (60%-85%)^{59,61,72}. The Phase 2 MEDIOLA trial of olaparib with durvalumab for patients with gBRCA1/2-mutated metastatic breast cancer reported an ORR of 63%, median PFS of 8.2 months, and median OS of 21.5 months²⁹⁷. A Phase 1 trial of olaparib with the PI3K inhibitor buparlisib reported an ORR of 33% (4/12) for patients with gBRCA1/2-mutated breast cancer²⁹⁸. A Phase 1 trial of olaparib plus carboplatin for patients with gBRCA1/2-mutated breast cancer reported an ORR of 88% (7/8)²⁹⁹. In a Phase 2 study of olaparib plus pembrolizumab for advanced solid tumors, patients with BRCA1 or BRCA2 mutations achieved an ORR of 29% (6/21), whereas patients with mutations in other homologous recombination repair genes achieved an ORR of 6.3% (2/32)³⁰⁰. Phase 1 trials of olaparib plus chemotherapy for patients with triple-negative breast cancer (TNBC) reported ORRs of 37-38%³⁰¹⁻³⁰². A small Phase 1 trial reported a 20% ORR (1/5) for patients with breast cancer and wild-type germline BRCA status following combination treatment with olaparib and buparlisib²⁹⁸. A Phase 2 study comparing durvalumab in combination with olaparib and paclitaxel to chemotherapy alone reported pathologic complete response (pCR) for 37% versus 22% of patients with HER2-negative breast cancer, 47% versus 27% of patients with TNBC, and 28% versus 14% of patients with HR-positive HER2-negative breast cancer³⁰³.

SAMPLE

THERAPIES WITH CLINICAL BENEFIT IN PATIENT'S TUMOR TYPE

ORDERED TEST #

Talazoparib

Assay findings association

BRCA1
V1234fs*8

AREAS OF THERAPEUTIC USE

The PARP inhibitor talazoparib is available in the EU as monotherapy to treat patients with HER2-negative locally advanced or metastatic breast cancer with germline BRCA mutations, who have been previously treated with, or are not considered candidates for, available therapies. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of strong clinical data in breast cancer^{80,304-305} and additional clinical evidence in ovarian, pancreatic, and prostate cancer³⁰⁶⁻³⁰⁹, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to talazoparib.

SUPPORTING DATA

In the Phase 3 EMBRACA trial, patients with HER2-negative advanced breast cancer and germline BRCA mutations achieved significantly longer median PFS (8.6 vs. 5.6 months, HR=0.54), a higher ORR (63% vs. 27%), and improved quality of life on talazoparib compared with standard chemotherapy (capecitabine, eribulin, gemcitabine, or vinorelbine)^{80,305}. Clinical benefit from talazoparib was observed for patients with either triple-negative or hormone receptor-positive (HR+) breast cancer, and for those with CNS metastases⁸⁰. Final OS analysis showed that talazoparib did not significantly improve OS compared with chemotherapy (median OS [mOS] 19.3 vs. 19.5 months, HR=0.85) but did significantly delay definitive clinically meaningful deterioration in global health status/quality of life³¹⁰. Retrospective

genomic analysis showed that MYC amplification was associated with significantly shorter mOS for patients with triple-negative cancer treated with talazoparib, but not for those treated with chemotherapy; in contrast, for patients with HR+ cancer, MYC amplification was associated with shorter mOS for the chemotherapy treatment group, but not for the talazoparib treatment group³¹¹. The efficacy of single-agent talazoparib for the treatment of BRCA-mutated advanced breast cancer was also demonstrated in earlier-phase studies, which reported ORRs of 21%-50%^{304,307}. As neoadjuvant treatment for BRCA-mutated HER2-negative breast cancer, talazoparib led to a pathologic complete response (pCR) for 53% (10/19) of patients³¹². In the Phase 2 I-SPY2 trial, talazoparib with synergy-dosed irinotecan (TI) for patients with early stage, high-risk HER2-negative breast cancer reported fewer Grade 3/4 adverse events compared with the chemotherapy control arm (paclitaxel with doxorubicin and cyclophosphamide [AC]), although a similar pCR rate was observed³¹³. Notably, 6/10 patients with germline BRCA mutations achieved a pCR with TI treatment³¹³. In a Phase 2 study of talazoparib for BRCA1/2-wildtype patients with homologous recombination pathway alterations, those with HER2-negative advanced breast cancer experienced an ORR of 31% (4/13 PRs), with responses observed for 3 patients with germline PALB2 mutations and for 1 patient with germline CHEK2 and FANCA mutations as well as somatic PTEN mutation; 3 additional patients with germline PALB2 or somatic ATR or PTEN alterations had SD ≥6 months³¹⁴.

SAMPLE

THERAPIES WITH CLINICAL BENEFIT | IN OTHER TUMOR TYPE

ORDERED TEST #

Niraparib

Assay findings association

BRCA1
V1234fs*8

AREAS OF THERAPEUTIC USE

The PARP inhibitor niraparib is available in the EU as a monotherapy to treat patients with epithelial high-grade ovarian, Fallopian tube, or peritoneal cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence in ovarian and breast cancers^{63-64,315}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to PARP inhibitors such as niraparib.

SUPPORTING DATA

In a Phase 1 study of niraparib treatment for patients with solid tumors, 2/4 patients with breast cancer and BRCA1/2 mutations experienced a PR⁶⁴. An open label study combining PD-1 inhibitor pembrolizumab with niraparib for patients with TNBC reported an ORR of 21% and DCR of 49%; ORR and DCR for patients with BRCA alterations were 47% and 80%, respectively, with 2 CRs, 5 PRs, 5 SDs and mPFS of 8.3 months³¹⁶.

Rucaparib

Assay findings association

BRCA1
V1234fs*8

AREAS OF THERAPEUTIC USE

The PARP inhibitor rucaparib is available in the EU to treat patients with platinum-sensitive relapsed or progressive BRCA mutated (germline and/or somatic) high-grade epithelial ovarian, fallopian tube, or primary peritoneal cancer who have been treated with 2 or more prior lines of platinum-based chemotherapy and who are unable to tolerate further platinum-based chemotherapy. Rucaparib is also available for the maintenance treatment of patients with platinum sensitive relapsed high-grade epithelial ovarian, fallopian tube, or primary peritoneal cancer who are in complete or partial response to platinum-based chemotherapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of strong clinical evidence in ovarian

cancer^{65-66,250}, as well as clinical data in other cancer types^{66,317-318}, loss or inactivation of either BRCA1 or BRCA2 may confer sensitivity to rucaparib.

SUPPORTING DATA

In a Phase 2 study evaluating rucaparib for patients with advanced breast or ovarian cancer and BRCA1/2 mutations, no objective responses were reported in breast cancer patients⁶⁶. However, 39% (9/23) of evaluable patients with breast cancer achieved stable disease lasting 12 weeks or more⁶⁶. In a Phase 1 study of rucaparib treatment in patients with solid tumors, 1 patient with breast cancer and a BRCA mutation given the recommended Phase 2 dose reported an objective response³¹⁷.

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies listed in this report may not be complete and exhaustive and the therapeutic agents are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type.

CLINICAL TRIALS

ORDERED TEST #

NOTE Clinical trials are ordered by gene and prioritized in the following descending order: Pediatric trial qualification → Geographical proximity → Trial phase → Trial verification within last 2 months. While every effort is made to ensure the accuracy of the information contained below, the

information available in the public domain is continually updated and should be investigated by the physician or research staff. The clinical trials listed in this report may not be complete and exhaustive or may include trials for which the patient does not meet the clinical trial

enrollment criteria. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov or local registries in your region.

GENE
BRCA1

RATIONALE
BRCA1 loss or inactivating alterations may predict sensitivity to PARP inhibitors or ATR inhibitors.

ALTERATION
V1234fs*8

NCT04915755

PHASE 3

Efficacy and Safety Comparison of Niraparib to Placebo in Participants With Either Human Epidermal Growth Factor 2 Negative (HER2-) Breast Cancer Susceptibility Gene Mutation (BRCAmut) or Triple-Negative Breast Cancer (TNBC) With Molecular Disease

TARGETS
PARP

LOCATIONS: Rzeszow (Poland), Krakow (Poland), Brindisi (Italy), Warszawa (Poland), Wroclaw (Poland), Napoli (Italy), Poznan (Poland), Meldola (FC) (Italy), Padova (Italy), Bologna (Italy)

NCT04768296

PHASE 2

Berzosertib + Topotecan in Relapsed Platinum-Resistant Small-Cell Lung Cancer (DDRiver SCLC 250)

TARGETS
TOP1, ATR

LOCATIONS: Meldola (Italy), Rome (Italy), Roma (Italy), Pisa (Italy), Milano (Italy), Strasbourg (France), Arlon (Belgium), Yvoir (Belgium), Brussels (Belgium), Gent (Belgium)

NCT04497116

PHASE 1/2

Study of RP-3500 in Advanced Solid Tumors

TARGETS
ATR, PARP

LOCATIONS: Copenhagen (Denmark), London (United Kingdom), Manchester (United Kingdom), Newcastle Upon Tyne (United Kingdom), Massachusetts, Rhode Island, New York, Toronto (Canada), North Carolina, Illinois

NCT03127215

PHASE 2

Study of Olaparib/Trabectedin vs. Doctor's Choice in Solid Tumors

TARGETS
FUS-DDIT3, PARP

LOCATIONS: Dresden (Germany), München (Germany), Stuttgart (Germany), Tuebingen (Germany), Heidelberg (Germany), Frankfurt (Germany), Freiburg (Germany), Mainz (Germany), Essen (Germany)

NCT04095273

PHASE 1

Study to Test How Well Patients With Advanced Solid Tumors Respond to Treatment With the ATR Inhibitor BAY1895344 in Combination With Pembrolizumab, to Find the Optimal Dose for Patients, How the Drug is Tolerated and the Way the Body Absorbs, Distributes and Discharges the Drug

TARGETS
ATR, PD-1

LOCATIONS: St. Gallen (Switzerland), Bellinzona (Switzerland), Tübingen (Germany), Heidelberg (Germany), Sutton (United Kingdom), Newcastle Upon Tyne (United Kingdom), Madrid (Spain), Massachusetts, Connecticut, New York

ORDERED TEST #

NCT02810743	PHASE 3
Substantially Improving the Cure Rate of High-risk BRCA1-like Breast Cancer	TARGETS PARP
LOCATIONS: Enschede (Netherlands), Marseille (France), Maastricht (Netherlands), Nijmegen (Netherlands), Groningen (Netherlands), Utrecht (Netherlands), Amsterdam (Netherlands), Rotterdam (Netherlands), Leiden (Netherlands)	
NCT02264678	PHASE 1/2
Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents	TARGETS ATR, PARP, PD-L1
LOCATIONS: Villejuif (France), Cambridge (United Kingdom), Sutton (United Kingdom), Bordeaux (France), Oxford (United Kingdom), Coventry (United Kingdom), Manchester (United Kingdom), Withington (United Kingdom), Massachusetts, New York	
NCT03907969	PHASE 1/2
A Clinical Trial to Evaluate AZD7648 Alone and in Combination With Other Anti-cancer Agents in Patients With Advanced Cancers	TARGETS PARP, DNA-PK
LOCATIONS: London (United Kingdom), Newcastle upon Tyne (United Kingdom), Connecticut, Texas	
NCT03150576	PHASE 2/3
Platinum and Polyadenosine 5'Diphosphoribose Polymerisation (PARP) Inhibitor for Neoadjuvant Treatment of Triple Negative Breast Cancer (TNBC) and/or Germline BRCA (gBRCA) Positive Breast Cancer	TARGETS PARP
LOCATIONS: Cambridge (United Kingdom)	
NCT04657068	PHASE 1/2
A Study of ART0380 for the Treatment of Advanced or Metastatic Solid Tumors	TARGETS ATR
LOCATIONS: London (United Kingdom), Pennsylvania, Tennessee, Florida, Colorado, Oklahoma	

ORDERED TEST #

GENE
PTEN

ALTERATION
loss

RATIONALE
PTEN loss or inactivating mutations may lead to increased activation of the PI₃K-AKT-mTOR pathway and may indicate sensitivity to inhibitors

of this pathway. PTEN loss or inactivation may also predict sensitivity to PARP inhibitors.

NCT03997123

PHASE 3

Capivasertib+Paclitaxel as First Line Treatment for Patients With Locally Advanced or Metastatic TNBC

TARGETS
AKTs

LOCATIONS: Košice (Slovakia), Prešov (Slovakia), Ankara (Turkey), Zilina (Slovakia), Bratislava (Slovakia), Heraklion (Greece), Guimarães (Portugal), Chengdu (China), Xi'an (China), Beijing (China)

NCT04915755

PHASE 3

Efficacy and Safety Comparison of Niraparib to Placebo in Participants With Either Human Epidermal Growth Factor 2 Negative (HER2-) Breast Cancer Susceptibility Gene Mutation (BRCAmut) or Triple-Negative Breast Cancer (TNBC) With Molecular Disease

TARGETS
PARP

LOCATIONS: Rzeszow (Poland), Krakow (Poland), Brindisi (Italy), Warszawa (Poland), Wroclaw (Poland), Napoli (Italy), Poznan (Poland), Meldola (FC) (Italy), Padova (Italy), Bologna (Italy)

NCT04862663

PHASE 3

Capivasertib + Palbociclib + Fulvestrant for HR+/HER2- Advanced Breast Cancer (CAPitello-292).

TARGETS
AKTs, CDK6, ER, CDK4

LOCATIONS: Kraków (Poland), Warszawa (Poland), Bydgoszcz (Poland), Odense C (Denmark), Solna (Sweden), Leuven (Belgium), Villejuif (France), Chicoutimi (Canada), St Herblain (France), Tennessee

NCT04770246

PHASE 2

TAS-117 in Patients With Advanced Solid Tumors Harboring Germline PTEN Mutations

TARGETS
AKT2, AKT1, AKT3

LOCATIONS: Vienna (Austria), Villejuif (France), London (United Kingdom), Pennsylvania, Ohio, Texas, California

NCT04802759

PHASE 1/2

A Study Evaluating the Efficacy and Safety of Multiple Treatment Combinations in Participants With Breast Cancer

TARGETS
ER, CDK4, CDK6, AKTs, PI3K-alpha, mTOR

LOCATIONS: Tel Aviv (Israel), Petach Tikva (Israel), Ramat Gan (Israel), Jerusalem (Israel), Barcelona (Spain), Valencia (Spain), Madrid (Spain), Massachusetts, Goyang-si (Korea, Republic of), Seoul (Korea, Republic of)

NCT04497116

PHASE 1/2

Study of RP-3500 in Advanced Solid Tumors

TARGETS
ATR, PARP

LOCATIONS: Copenhagen (Denmark), London (United Kingdom), Manchester (United Kingdom), Newcastle Upon Tyne (United Kingdom), Massachusetts, Rhode Island, New York, Toronto (Canada), North Carolina, Illinois

CLINICAL TRIALS

ORDERED TEST #

NCT02264678

PHASE 1/2

Ascending Doses of AZD6738 in Combination With Chemotherapy and/or Novel Anti Cancer Agents

TARGETS
ATR, PARP, PD-L1

LOCATIONS: Villejuif (France), Cambridge (United Kingdom), Sutton (United Kingdom), Bordeaux (France), Oxford (United Kingdom), Coventry (United Kingdom), Manchester (United Kingdom), Withington (United Kingdom), Massachusetts, New York

NCT04464174

PHASE 2

Ipatasertib Plus Non-Taxane Chemotherapy for Advanced or Metastatic Triple-Negative Breast Cancer

TARGETS
AKTs

LOCATIONS: Barcelona (Spain), Lleida (Spain), Castellón De La Plana (Spain), Zaragoza (Spain), Valencia (Spain), Alicante (Spain), El Palmar (Spain), Madrid (Spain), Málaga (Spain), Cáceres (Spain)

NCT03907969

PHASE 1/2

A Clinical Trial to Evaluate AZD7648 Alone and in Combination With Other Anti-cancer Agents in Patients With Advanced Cancers

TARGETS
PARP, DNA-PK

LOCATIONS: London (United Kingdom), Newcastle upon Tyne (United Kingdom), Connecticut, Texas

NCT03150576

PHASE 2/3

Platinum and Polyadenosine 5'Diphosphoribose Polymerisation (PARP) Inhibitor for Neoadjuvant Treatment of Triple Negative Breast Cancer (TNBC) and/or Germline BRCA (gBRCA) Positive Breast Cancer

TARGETS
PARP

LOCATIONS: Cambridge (United Kingdom)

ORDERED TEST #

APPENDIX Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

EPHA3
D344V

ESR1
S118P

MLL2
Q3745_H3746insQ

NF2
rearrangement

POLE
R2165C

PRDM1
amplification

RAD51D
I311N

STAG2
H774N

SAMPLE

ORDERED TEST #

APPENDIX

Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B or WTX)	
APC	AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX
AURKA	AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2
BCL6	BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1
BTG2	BTK	CALR	CARD11	CASP8	CBFB	CBL	CCND1	CCND2
CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73	CDH1
CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B	CDKN2C
CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R	CTCF
CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1	DDR2
DIS3	DNMT3A	DOT1L	EED	EGFR	EMSY (C11orf30)	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14
FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3	FGFR4
FH	FLCN	FLT1	FLT3	FOXL2	FUBP1	GABRA6	GATA3	GATA4
GATA6	GID4 (C17orf39)	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3-3A (H3F3A)
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMSA	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11 (MRE11A)	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD2 (WHSC1 or MMSET)		NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3
P2RY8	PALB2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)
PDGFRA	PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1
PMS2	POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI
PRKN (PARK2)	PTCH1	PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51
RADS1B	RADS1C	RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10
REL	RET	RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO
SNCAIP	SOC1	SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3
STK11	SUFU	SYK	TBX3	TEK	TENTSC (FAM46C)	TET2	TGFBR2	TIPARP
TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL
WT1	XPO1	XRCC2	ZNF217	ZNF703				

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV1
ETV4	ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT
KMT2A (MLL)	MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA
RAF1	RARA	RET	ROS1	RSPO2	SDC4	SLC34A2	TERC*	TERT**
TMPPRSS2								

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER GENOMIC SIGNATURES

- Loss of Heterozygosity (LOH) score
- Microsatellite (MS) status
- Tumor Mutational Burden (TMB)

ORDERED TEST #

APPENDIX
About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Ciplastraat 3, 2440 Geel, Belgium.



ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes. Using an Illumina® HiSeq platform, hybrid

capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: The association of a therapy with a genomic alteration or signature does not necessarily indicate pharmacologic effectiveness (or lack thereof); no association of a therapy with a genomic alteration or signature does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness).

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss - equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Genomic signatures and gene alterations detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each genomic signature or gene alteration. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X,

© 2022 Foundation Medicine, Inc. All rights reserved.

ORDERED TEST #

APPENDIX

About FoundationOne®CDx

- “MS-Equivocal,” or “Cannot Be Determined” should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI’s TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as “Cannot Be Determined” if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
 - Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient’s physician should determine whether the patient is a candidate for biopsy.
 - Reflex testing to an alternative FDA approved

companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient’s tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively

ORDERED TEST #

APPENDIX

About FoundationOne®CDx

determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

REFERENCE SEQUENCE INFORMATION

Sequence data is mapped to the human genome, Genome Reference Consortium Human Build 37 (GRCh37), also known as hg19.

MR Suite Version 6.2.0

The median exon coverage for this sample is 1,247x

ORDERED TEST #

APPENDIX **References**

157. Chen SH, et al. *Oncogene* (2018) PMID: 29059158
 158. Hennessy BT, et al. *Cancer Res.* (2009) PMID: 19435916
 159. Mercapide J, et al. *Mol. Carcinog.* (2002) PMID: 12203362
 160. Hohensee I, et al. *Am. J. Pathol.* (2013) PMID: 23665199
 161. Perez EA, et al. *J. Clin. Oncol.* (2013) PMID: 23650412
 162. Tsutsui S, et al. *Oncology* (2005) PMID: 16020969
 163. Zhang HY, et al. *Oncol Lett* (2013) PMID: 23946797
 164. Capodanno A, et al. *Hum. Pathol.* (2009) PMID: 19428048
 165. Campbell RB, et al. *J. Biol. Chem.* (2003) PMID: 12857747
 166. Rodríguez-Escudero I, et al. *Hum. Mol. Genet.* (2011) PMID: 21828076
 167. He X, et al. *Cancer Res.* (2013) PMID: 23475934
 168. Han SY, et al. *Cancer Res.* (2000) PMID: 10866302
 169. Myers MP, et al. *Proc. Natl. Acad. Sci. U.S.A.* (1998) PMID: 9811831
 170. Pradella LM, et al. *BMC Cancer* (2014) PMID: 24498881
 171. Kim JS, et al. *Mol. Cell. Biol.* (2011) PMID: 21536651
 172. Denning G, et al. *Oncogene* (2007) PMID: 17213812
 173. Hlobilkova A, et al. *Anticancer Res.* () PMID: 16619501
 174. Redfern RE, et al. *Protein Sci.* (2010) PMID: 20718038
 175. Shenoy S, et al. *PLoS ONE* (2012) PMID: 22505997
 176. Wang Y, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2009) PMID: 19329485
 177. Okumura K, et al. *J. Biol. Chem.* (2006) PMID: 16829519
 178. Lee JO, et al. *Cell* (1999) PMID: 10555148
 179. Maxwell GL, et al. *Cancer Res.* (1998) PMID: 9635567
 180. Risinger JI, et al. *Clin. Cancer Res.* (1998) PMID: 9865913
 181. Kato H, et al. *Clin. Cancer Res.* (2000) PMID: 11051241
 182. Fenton TR, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2012) PMID: 22891331
 183. Ngeow J, et al. *J. Clin. Endocrinol. Metab.* (2012) PMID: 23066114
 184. Lobo GP, et al. *Hum. Mol. Genet.* (2009) PMID: 19457929
 185. Liu J, et al. *Oncogene* (2014) PMID: 23995781
 186. Maehama T, et al. *Annu. Rev. Biochem.* (2001) PMID: 11395408
 187. De Vivo I, et al. *J. Med. Genet.* (2000) PMID: 10807691
 188. Ramaswamy S, et al. *Proc. Natl. Acad. Sci. U.S.A.* (1999) PMID: 10051603
 189. Liu JL, et al. *Mol. Cell. Biol.* (2005) PMID: 15988030
 190. Karoui M, et al. *Br. J. Cancer* (2004) PMID: 15026806
 191. Gil A, et al. *PLoS ONE* (2015) PMID: 25875300
 192. Furnari FB, et al. *Cancer Res.* (1998) PMID: 9823298
 193. Spinelli L, et al. *J. Med. Genet.* (2015) PMID: 25527629
 194. Mingo J, et al. *Eur. J. Hum. Genet.* (2018) PMID: 29706633
 195. Wang Q, et al. *J. Mol. Graph. Model.* (2010) PMID: 20538496
 196. Andrés-Pons A, et al. *Cancer Res.* (2007) PMID: 17942903
 197. Butler MG, et al. *J. Med. Genet.* (2005) PMID: 15805158
 198. Georgescu MM, et al. *Proc. Natl. Acad. Sci. U.S.A.* (1999) PMID: 10468583
 199. Staal FJ, et al. *Br. J. Cancer* (2002) PMID: 12085208
 200. Nguyen HN, et al. *Oncogene* (2014) PMID: 24292679
 201. Rahdar M, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2009) PMID: 19114656
 202. Das S, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2003) PMID: 12808147
 203. Wang X, et al. *Biochem. J.* (2008) PMID: 18498243
 204. Valiente M, et al. *J. Biol. Chem.* (2005) PMID: 15951562
 205. Nguyen HN, et al. *Oncogene* (2015) PMID: 25263454
 206. Shan L, et al. *Cell Discov* (2020) PMID: 32704382
 207. Blumenthal GM, et al. *Eur. J. Hum. Genet.* (2008) PMID: 18781191
 208. Orloff MS, et al. *Oncogene* (2008) PMID: 18794875
 209. Zbuk KM, et al. *Nat. Rev. Cancer* (2007) PMID: 17167516
 210. Zauderer et al., 2018; ASCO Abstract 8515
 211. LaFave LM, et al. *Nat. Med.* (2015) PMID: 26437366
 212. Yu H, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2014) PMID: 24347639
 213. Ismail IH, et al. *Cancer Res.* (2014) PMID: 24894717
 214. Peña-Llopis S, et al. *Nat. Genet.* (2012) PMID: 22683710
 215. Nishi R, et al. *Nat. Cell Biol.* (2014) PMID: 25194926
 216. Landreville S, et al. *Clin. Cancer Res.* (2012) PMID: 22038994
 217. Mendes-Pereira AM, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2012) PMID: 21482774
 218. Je EM, et al. *APMIS* (2012) PMID: 22958294
 219. Stephens PJ, et al. *Nature* (2012) PMID: 22722201
 220. Shah SP, et al. *Nature* (2012) PMID: 22495314
 221. Zarrizi R, et al. *Cancer Res.* (2014) PMID: 25228651
 222. Jensen DE, et al. *Oncogene* (1998) PMID: 9528852
 223. Ventii KH, et al. *Cancer Res.* (2008) PMID: 18757409
 224. Chan-On W, et al. *Nat. Genet.* (2013) PMID: 24185513
 225. Abdel-Rahman MH, et al. *J. Med. Genet.* (2011) PMID: 21941004
 226. Testa JR, et al. *Nat. Genet.* (2011) PMID: 21874000
 227. Wiesner T, et al. *Nat. Genet.* (2011) PMID: 21874003
 228. Farley MN, et al. *Mol. Cancer Res.* (2013) PMID: 23709298
 229. Aoude LG, et al. *PLoS ONE* (2013) PMID: 23977234
 230. Peng H, et al. *Cancer Res* (2018) PMID: 29284740
 231. Nishikawa H, et al. *Cancer Res* (2009) PMID: 19117993
 232. Zhang Y, et al. *Nat Cell Biol* (2018) PMID: 30202049
 233. Walpole S, et al. *J Natl Cancer Inst* (2018) PMID: 30517737
 234. Boru G, et al. *Genes Chromosomes Cancer* (2019) PMID: 30883995
 235. Garfield EM, et al. *J Am Acad Dermatol* (2018) PMID: 29753057
 236. Shankar GM, et al. *Neuro Oncol* (2017) PMID: 28170043
 237. Rai K, et al. *Genes Chromosomes Cancer* (2017) PMID: 27178540
 238. Zauderer MG, et al. *J Thorac Oncol* (2019) PMID: 31323388
 239. Hirai H, et al. *Cancer Biol. Ther.* (2010) PMID: 20107315
 240. Bridges KA, et al. *Clin. Cancer Res.* (2011) PMID: 21799033
 241. Rajeshkumar NV, et al. *Clin. Cancer Res.* (2011) PMID: 21389100
 242. Osman AA, et al. *Mol. Cancer Ther.* (2015) PMID: 25504633
 243. Xu L, et al. *Mol. Cancer Ther.* (2002) PMID: 12489850
 244. Xu L, et al. *Mol. Med.* (2001) PMID: 11713371
 245. Camp ER, et al. *Cancer Gene Ther.* (2013) PMID: 23470564
 246. Kim SS, et al. *Nanomedicine* (2015) PMID: 25240597
 247. Pirolo KF, et al. *Mol. Ther.* (2016) PMID: 27357628
 248. Hajdenberg et al., 2012; ASCO Abstract e15010
 249. Leijen S, et al. *J. Clin. Oncol.* (2016) PMID: 27601554
 250. Moore et al., 2019; ASCO Abstract 5513
 251. Leijen S, et al. *J. Clin. Oncol.* (2016) PMID: 27998224
 252. Oza et al., 2015; ASCO Abstract 5506
 253. Lee J, et al. *Cancer Discov* (2019) PMID: 31315834
 254. Méndez E, et al. *Clin. Cancer Res.* (2018) PMID: 29535125
 255. Seligmann JF, et al. *J Clin Oncol* (2021) PMID: 34538072
 256. Lehmann S, et al. *J. Clin. Oncol.* (2012) PMID: 22965953
 257. Mohell N, et al. *Cell Death Dis* (2015) PMID: 26086967
 258. Fransson Å, et al. *J Ovarian Res* (2016) PMID: 27179933
 259. Gourley et al., 2016; ASCO Abstract 5571
 260. Kwok M, et al. *Blood* (2016) PMID: 26563132
 261. Boudny M, et al. *Haematologica* (2019) PMID: 30975914
 262. Dillon MT, et al. *Mol. Cancer Ther.* (2017) PMID: 28062704
 263. Middleton FK, et al. *Cancers (Basel)* (2018) PMID: 30127241
 264. Banerji S, et al. *Nature* (2012) PMID: 22722202
 265. Alsner J, et al. *Acta Oncol* (2008) PMID: 18465328
 266. Alkam Y, et al. *Histopathology* (2013) PMID: 24004112
 267. Uji K, et al. *Cancer Lett.* (2014) PMID: 23973262
 268. Olivier M, et al. *Clin. Cancer Res.* (2006) PMID: 16489069
 269. Végan F, et al. *PLoS ONE* (2013) PMID: 23359294
 270. Walsh T, et al. *JAMA* (2006) PMID: 16551709
 271. Garber JE, et al. *J. Clin. Oncol.* (2005) PMID: 15637391
 272. Apostolou P, et al. *Biomed Res Int* (2013) PMID: 23586058
 273. Brown CJ, et al. *Nat. Rev. Cancer* (2009) PMID: 19935675
 274. Joerger AC, et al. *Annu. Rev. Biochem.* (2008) PMID: 18410249
 275. Kato S, et al. *Proc. Natl. Acad. Sci. U.S.A.* (2003) PMID: 12826609
 276. Kamada R, et al. *J. Biol. Chem.* (2011) PMID: 20978130
 277. Zerdoumi Y, et al. *Hum. Mol. Genet.* (2017) PMID: 28472496
 278. Yamada H, et al. *Carcinogenesis* (2007) PMID: 17690113
 279. Bougeard G, et al. *J. Clin. Oncol.* (2015) PMID: 26014290
 280. Sorrell AD, et al. *Mol Diagn Ther* (2013) PMID: 23355100
 281. Nichols KE, et al. *Cancer Epidemiol. Biomarkers Prev.* (2001) PMID: 11219776
 282. Kleihues P, et al. *Am. J. Pathol.* (1997) PMID: 9006316
 283. Gonzalez KD, et al. *J. Clin. Oncol.* (2009) PMID: 19204208
 284. Lalloo F, et al. *Lancet* (2003) PMID: 12672316
 285. Mandelker D, et al. *Ann. Oncol.* (2019) PMID: 31050713
 286. Jaiswal S, et al. *N. Engl. J. Med.* (2014) PMID: 25426837
 287. Genovese G, et al. *N. Engl. J. Med.* (2014) PMID: 25426838
 288. Xie M, et al. *Nat. Med.* (2014) PMID: 25326804
 289. Acuna-Hidalgo R, et al. *Am. J. Hum. Genet.* (2017) PMID: 28669404
 290. Severson EA, et al. *Blood* (2018) PMID: 29678827
 291. Fuster JJ, et al. *Circ. Res.* (2018) PMID: 29420212
 292. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
 293. Chabon JJ, et al. *Nature* (2020) PMID: 32269342
 294. Razavi P, et al. *Nat. Med.* (2019) PMID: 31768066
 295. Del Conte G, et al. *Br. J. Cancer* (2014) PMID: 25025963
 296. Tutt et al., 2022; ESMO Plenary Abstract VPI-2022
 297. Domchek SM, et al. *Lancet Oncol* (2020) PMID: 32771088
 298. Matulonis UA, et al. *Ann. Oncol.* (2017) PMID: 27993796
 299. Lee JM, et al. *J. Natl. Cancer Inst.* (2014) PMID: 24842883
 300. Maio et al., 2021; AACR Abstract CT178
 301. Takahashi et al., 2016; ASCO Abstract 1080
 302. Dent RA, et al. *Breast Cancer Res.* (2013) PMID: 24063698
 303. Pusztaí et al., 2020; AACR Abstract CT011
 304. Turner et al., 2017; ASCO Abstract 1007
 305. Ettl J, et al. *Ann. Oncol.* (2018) PMID: 30124753
 306. Meehan et al., 2017; AACR Abstract 4687
 307. de Bono J, et al. *Cancer Discov* (2017) PMID: 28242752
 308. Lu E, et al. *J Natl Compr Canc Netw* (2018) PMID: 30099369

ORDERED TEST #

APPENDIX

References

309. De Bono et al., 2020; ASCO Abstract 5566
 310. Litton JK, et al. Ann Oncol (2020) pmid: 32828825
 311. Ettl et al., 2020; SABCS Abstract P55-07
 312. Litton JK, et al. J. Clin. Oncol. (2019) pmid: 31461380

313. Schwab et al., 2019; AACR Abstract CT123/2
 314. Gruber et al., 2019; ASCO Abstract 3006
 315. Konstantinopolous et al., 2018; ASCO Abstract 106
 316. Vinayak S, et al. JAMA Oncol (2019) pmid: 31194225

317. Kristeleit et al., 2014; ASCO Abstract 2573
 318. Domcheck et al., 2016; ASCO Abstract 4110

SAMPLE